

lactone,⁹ have been reported; in each case, the isolation procedure included exposure to high temperatures. In view of the demonstrated transformation of germacranolide precursors to elemanolides by heating, the question has been raised as to whether some of the elemanolides may be artifacts.^{8,9} In contrast, when an isolation procedure was devised which involved cold aqueous extraction of *V. hymenolepis*, vernolepin was isolated in a yield comparable to that obtained by hot ethanol extraction. This fact supports the view that vernolepin is, indeed, a naturally occurring compound.

Vernolepin and vernomenin appear to be the first recognized elemanolide dilactones.

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The Fate of the 15 β Hydrogen of Lanosterol in Cholesterol Biosynthesis

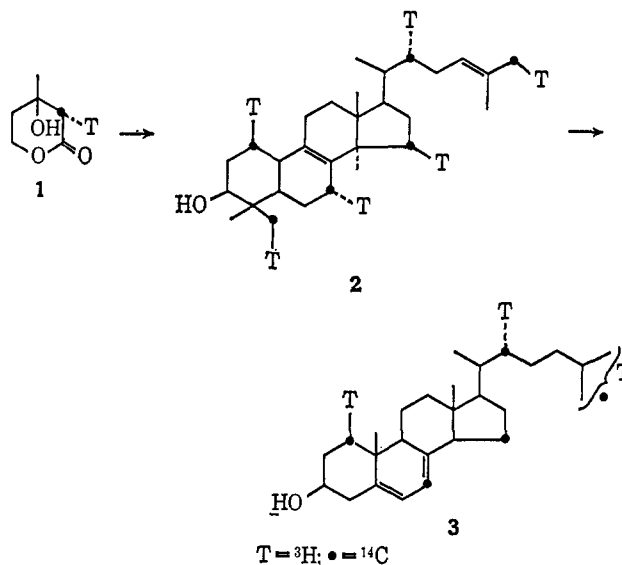
Sir:

In cholesterol biosynthesis the three methyl groups, 4 α , 4 β , and 14 α , of lanosterol are removed by oxidation to carbon dioxide.¹ The 14 α -methyl group is believed to be transformed into a carboxyl group and eliminated by decarboxylation, which is facilitated by the 8,9 double bond.^{2,3}

If the 8,9 double bond is not the only promoter of the elimination of the 14 α -carboxyl group, this reaction could be facilitated by one of the following mechanisms: (a) oxidation of 14 α -carboxylic acid with formation of a carboxyl radical and its elimination with the involvement of one of the hydrogen atoms in position 15, (b) oxidation in position 15 and concerted elimination of the carboxyl group, (c) previous formation of double bond in position 15,16 facilitating the elimination of carbon in 14. All these possibilities imply the removal of hydrogen atoms in position 15. The fate of these hydrogen atoms during cholesterol biosynthesis has been followed by determining the position of labeled hydrogens in lanosterol, 5 α -cholest-7-en-3 β -ol, and cholesta-5,7-dien-3 β -ol biosynthesized from labeled mevalonic acids. Actual ³H:¹⁴C ratios and atom equivalents of all the significant compounds are shown in Table I.

Accumulation of labeled cholesta-5,7-dien-3 β -ol (3) has been obtained by incubating liver homogenates⁴ of rats pretreated with AY 9944,⁵ in the presence of the

same inhibitor⁶ and of 3(\pm)-(2*S*)-[2-¹⁴C-2-³H]mevalonic acid lactone (1) (10 μ Ci of ¹⁴C, ³H:¹⁴C 10.00).⁷ Radioactive carbon atoms in cholesta-5,7-dien-3 β -ol correspond to the positions shown in formula 3.⁸ Since the radioactive precursor is asymmetrically labeled, tritium should be localized at positions 1 α , 15 β , 22*R*, and 26 or 27, the 7 position being excluded on the basis of our previous results.⁹ The ³H:¹⁴C ratio should be 4:5. If the 15 β hydrogen is exchanged with the medium, this ratio should be 3:5.



The unsaponifiable residue from homogenates was acetylated, carrier cholesta-5,7-dien-3 β -ol acetate was added, and the mixture was separated by column chromatography on silver nitrate-kieselgel G-Celite.¹⁰ The obtained cholesta-5,7-dien-3 β -ol acetate¹¹ was diluted again with nonradioactive material and hydrogenated in presence of tris(triphenylphosphine)rhodium chloride⁶ to yield 5 α -cholest-7-en-3 β -ol acetate.¹¹ After column chromatography on silver nitrate-kieselgel G-Celite,¹⁰ this compound (0.565 μ Ci of ¹⁴C/mmmole) showed a ³H:¹⁴C ratio of 6.07, corresponding to 3.02 labeled hydrogens out of 5 radioactive carbon atoms. The ¹⁴C radioactivity and the ³H:¹⁴C ratio were constant after several crystallizations. Furthermore oxidation with osmium tetroxide of some of the radioactive 5 α -cholest-7-en-3 β -ol acetate produced the mixture of the epimeric *cis*-5 α -cholestane-3 β ,7,8-triol 3 β -acetates^{6,11} (0.563 μ Ci of ¹⁴C/mmmole) which showed an unchanged ³H:¹⁴C ratio with respect to 5 α -cholest-7-en-3 β -ol acetate. The same ratio was found in the mixture of the epimeric *cis*-5 α -cholestane-3 β ,7,8-triol 3 β -acetates^{6,10} (0.189 μ Ci of ¹⁴C/mmmole) obtained from radioactive 5 α -cholest-7-en-3 β -ol acetate which could be isolated in small amounts⁶ from liver homogenates. The expected constant ³H:¹⁴C ratio was also found

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(7) Incubation experiments were performed at least in duplicate; reproducibility of results was excellent.

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(10) G. Galli and E. G. Paoletti, *Lipids*, **2**, 72 (1967); **2**, 84 (1967).

(11) The chemical purity of all compounds was established by comparing melting points, optical rotation values, mass spectra, and glpc retention times on a 1% phenylsilicone glass column with those of authentic samples.

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(2) F. Gautschi and K. Bloch, *J. Am. Chem. Soc.*, **79**, 684 (1957).

(3) M. Lindberg, F. Gautschi, and K. Bloch, *J. Biol. Chem.*, **238**, 1661 (1963).

(4) N. L. R. Bucher and K. McGarrah, *ibid.*, **222**, 1 (1956).

(5) *trans*-1,4-Bis(2-chlorobenzylaminomethyl)cyclohexane hydrochloride, a specific inhibitor of cholesta-5,7-dien-3 β -ol- Δ^7 -reductase, according to D. Dvornik, M. Kraml, J. Dubuc, M. Givner, and R. Gaudry, *J. Am. Chem. Soc.*, **85**, 3309 (1963).

in the mixture of the epimeric 5 α -cholestane-3 α ,8-diol-7-one 3 β -acetates¹¹ (0.564 μ Ci of ¹⁴C/mmmole) resulting from the mixture of the epimeric *cis*-5 α -cholestane-3 β ,7,8-triol 3 β -acetates.⁸

In order to establish if the missing tritium was originally from the 15 position, the remaining 5 α -cholest-7-en-3 β -ol acetate (0.565 μ Ci of ¹⁴C/mmmole) was oxidized with selenium dioxide¹² to 5 α -cholest-8(14)-en-3 β ,7 α -diol diacetate¹¹ which, adsorbed on silver nitrate-kieselgel G-Celite, gave rise to 5 α -cholesta-7,14-dien-3 β -ol acetate: mp 67–69°; uv max (C₂H₅OH) 242 m μ (ϵ 9700).¹³ This diene was oxidized with chromium trioxide in acetic acid to yield 5 α -cholest-8(14)-en-3 β -ol-7,15-dione acetate; mp 153–155°; uv max (C₂H₅OH) 260 m μ (ϵ 6200); [α]_D²⁵ 65 (c 1, CHCl₃)¹³ (0.567 μ Ci of ¹⁴C/mmmole). This compound showed a ³H:¹⁴C ratio identical with that of 5 α -cholest-7-en-3 β -ol acetate. The formation of a keto group in position 15, unaccompanied by a decrease of the ³H:¹⁴C ratio, shows that the 15 β hydrogen of **3** is not radioactive.

In order to establish the stereospecificity of the exchange of hydrogens in position 15, a biosynthetic experiment has been performed starting from 3(\pm)-(2*R*)-[2-¹⁴C-2-³H]mevalonic acid lactone (10 μ Ci of ¹⁴C, ³H:¹⁴C 9.20). Incubation and isolation procedures were the same as described above. Radioactive cholesta-5,7-dien-3 β -ol acetate¹¹ was transformed and purified as in the previous experiment, and the 5 α -cholest-7-en-3 β -ol acetate¹¹ obtained (0.581 μ Ci of ¹⁴C/mmmole) showed a ³H:¹⁴C ratio 8.97 corresponding to 4.90 labeled hydrogens out of 5 radioactive carbon atoms. This result shows that the exchange of the 15 β hydrogen is stereospecific.

The loss of hydrogen from the 15 β position has been followed in cholesterol biosynthesis after the cyclization of squalene to lanosterol. Accumulation of labeled lanosterol (**2**) has been obtained by incubating rat liver homogenates with 3(\pm)-(2*S*)-[2-¹⁴C-2-³H]mevalonic acid lactone (10 μ Ci of ¹⁴C, ³H:¹⁴C 10.16) in presence of 10⁻³ M sodium arsenite.¹⁴ The unsaponifiable residue was acetylated, carrier lanosterol acetate (lanosta-8,24-dien-3 β -ol acetate) was added, and the mixture was separated by tlc¹⁵ on kieselgel G impregnated with silver nitrate. The isolated lanosterol acetate¹¹ showed a ³H:¹⁴C ratio corresponding to 5.74 labeled hydrogens out of 6 radioactive carbon atoms. This rather low ratio could have been caused by contamination of small amounts of demethyl analogs of lanosterol which are difficult to separate. Chromic acid oxidation¹⁶ of radioactive lanosterol acetate gave rise to methyl 25,26,27-trisnor-3 β -acetoxy lanost-8-en-7,11-dione-24-oate¹¹ with a ³H:¹⁴C ratio corresponding to 3.95 labeled hydrogens out of 5 radioactive carbon atoms. This result proves that the lanosterol retains the radioactive 15 β hydrogen.

It seems unlikely that the expulsion of the 4,4-dimethyl groups occurs with involvement of the 15 β hydrogen. However the isomerization of 5 α -cholest-8-en-3 β -ol into 5 α -cholest-7-en-3 β -ol is proved to proceed through incorporation of only one hydrogen atom from

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(13) All melting points are uncorrected. Satisfactory elementary analyses were obtained for all new compounds.

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Table I

Compound	³ H: ¹⁴ C, μ Ci: μ Ci	Atom equiv	
		Found	Calcd
3(\pm)-(2 <i>S</i>)-[2- ¹⁴ C-2- ³ H]Mevalonic acid lactone	10.00		
5 α -Cholest-7-en-3 β -ol acetate	6.07	3.02/5	3/5
<i>cis</i> -5 α -Cholestane-3 β ,7,8-triol 3 β -acetate mixture	6.07	3.02/5	3/5
5 α -Cholestane-3 β ,8-diol-7-one 3 β -acetate	6.05	3.01/5	3/5
5 α -Cholest-8(14)-en-3 β -ol-7,15-dione acetate	5.90	2.94/5	3/5
3(\pm)-(2 <i>S</i>)-[2- ¹⁴ C-2- ³ H]Mevalonic acid lactone	10.16		
Lanosta-8,24-dien-3 β -ol acetate	9.70	5.74/6	6/6
25,26,27-Trisnor-3 β -acetoxy lanost-8-en-7,11-dion-24-oate	6.68	3.95/5	4/5
3(\pm)-(2 <i>R</i>)-[2- ¹⁴ C-2- ³ H]Mevalonic acid lactone	9.20		
5 α -Cholest-7-en-3 β -ol acetate	8.97	4.90/5	5/5

the medium¹⁷ and it is not intramolecular in nature.⁸ Therefore it seems probable that the elimination of the 15 β hydrogen accompanies the expulsion of the 14 α -methyl group.

Further aspects of this elimination are under investigation.

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Halonium Ylides. I

Sir:

We wish to advocate the existence of bromonium ylides as reaction intermediates and to present preliminary findings concerning the chemistry of such ylides. Irradiation (>4800 Å, ~5–15°) of a degassed hexafluorobenzene solution 0.19 M in 3,5-di-*t*-butylbenzene 1,4-diazooxide¹ (**1**) and 0.94 M in 2,6-diisopropyl-4-bromophenol (**2**) leads to the precipitation of diaryl-bromonium bromide **3** as a reddish syrup isolable in ca. 15% yield after crystallization from hexafluorobenzene, *n*-hexane, and methylene chloride mixtures. The colorless salt has an elemental composition (*Anal.*

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